TABLE 6—ACID EXTRACTABLE COMPOUND CHARACTERISTIC m/z’S

<table>
<thead>
<tr>
<th>Compound</th>
<th>Labeled Analog</th>
<th>Primary m/z 1</th>
</tr>
</thead>
<tbody>
<tr>
<td>p-cresol 2</td>
<td></td>
<td>108/116</td>
</tr>
</tbody>
</table>

m/z = mass to charge ratio.
1 Native/labeled.
2 Analysis of this pollutant is approved only for the Centralized Waste Treatment industry.
3 Analysis of this pollutant is approved only for the Centralized Waste Treatment and Landfills industries.

TABLE 7—ACCEPTANCE CRITERIA FOR PERFORMANCE TESTS

<table>
<thead>
<tr>
<th>EGD No.</th>
<th>Compound</th>
<th>Acceptance criteria</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Initial precision and accuracy section 8.2 (μg/L)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Labeled compound recovery sec. 8.3 and 14.2 P (percent)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Calibration verification sec. 12.5 (μg/mL)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>On-going accuracy sec. 12.7 R (μg/L)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>S X s</td>
</tr>
<tr>
<td>758</td>
<td>acetophenone 1</td>
<td>34 44–167</td>
</tr>
<tr>
<td>658</td>
<td>acetophenone-d 1</td>
<td>51 23–254</td>
</tr>
<tr>
<td>757</td>
<td>aniline 1</td>
<td>32 30–171</td>
</tr>
<tr>
<td>657</td>
<td>aniline-d 2</td>
<td>71 15–327</td>
</tr>
<tr>
<td>771</td>
<td>o-cresol 1</td>
<td>40 31–326</td>
</tr>
<tr>
<td>1744</td>
<td>p-cresol 2</td>
<td>23 30–146</td>
</tr>
<tr>
<td>1644</td>
<td>p-cresol-d 2</td>
<td>59 54–140</td>
</tr>
<tr>
<td>757</td>
<td>aniline 1</td>
<td>22 11–318</td>
</tr>
<tr>
<td>671</td>
<td>aniline-d 2</td>
<td>13 40–160</td>
</tr>
<tr>
<td>1330</td>
<td>pyridine 1</td>
<td>28 10–421</td>
</tr>
<tr>
<td>1230</td>
<td>pyridine-d 2</td>
<td>ns 7–392</td>
</tr>
</tbody>
</table>

s = Standard deviation of four recovery measurements.
X = Average recovery for four recovery measurements.
EGD = Effluent Guidelines Division.
ns = no specification; limit is outside the range that can be measured reliably.
1 Analysis of this pollutant is approved only for the Centralized Waste Treatment industry.
2 Analysis of this pollutant is approved only for the Centralized Waste Treatment and Landfills industries.

APPENDIX B TO PART 136—DEFINITION AND PROCEDURE FOR THE DETERMINATION OF THE METHOD DETECTION LIMIT—REVISION 1.11

**Definition**

The method detection limit (MDL) is defined as the minimum concentration of a substance that can be measured and reported with 99% confidence that the analyte concentration is greater than zero and is determined from analysis of a sample in a given matrix containing the analyte.

**Scope and Application**

This procedure is designed for applicability to a wide variety of sample types ranging from reagent (blank) water containing analyte to wastewater containing analyte. The MDL for an analytical procedure may vary as a function of sample type. The procedure requires a complete, specific, and well defined analytical method. It is essential that all sample processing steps of the analytical method be included in the determination of the method detection limit.

The MDL obtained by this procedure is used to judge the significance of a single measurement of a future sample. The MDL procedure was designed for applicability to a broad variety of physical and chemical methods. To accomplish this, the procedure was made device- or instrument-independent.

**Procedure**

1. Make an estimate of the detection limit using one of the following:
   (a) The concentration value that corresponds to an instrument signal/noise in the range of 2.5 to 5.
   (b) The concentration equivalent of three times the standard deviation of replicate instrumental measurements of the analyte in reagent water.
   (c) That region of the standard curve where there is a significant change in sensitivity, i.e., a break in the slope of the standard curve.
(d) Instrumental limitations.

It is recognized that the experience of the analyst is important to this process. However, the analyst must include the above considerations in the initial estimate of the detection limit.

2. Prepare reagent (blank) water that is as free of analyte as possible. Reagent or interference free water is defined as a water sample in which analyte and interferent concentrations are not detected at the method detection limit of each analyte of interest. Interferences are defined as systematic errors in the measured analytical signal of an established procedure caused by the presence of interfering species (interferent). The interferent concentration is presupposed to be normally distributed in representative samples of a given matrix.

3. (a) If the MDL is to be determined in reagent (blank) water, prepare a laboratory standard (analyte in reagent water) at a concentration which is at least equal to or in the same concentration range as the estimated method detection limit. (Recommend between 1 and 5 times the estimated method detection limit.) Proceed to Step 4.

(b) If the MDL is to be determined in another sample matrix, analyze the sample. If the measured level of the analyte is in the recommended range of one to five times the estimated detection limit, proceed to Step 4. If the measured level of analyte is less than the estimated detection limit, add a known amount of analyte to bring the level of analyte between one and five times the estimated detection limit. If the measured level of analyte is greater than five times the estimated detection limit, there are two options.

(1) Obtain another sample with a lower level of analyte in the same matrix if possible.

(2) The sample may be used as is for determining the method detection limit if the analyte level does not exceed 10 times the MDL of the analyte in reagent water. The variance of the analytical method changes as the analyte concentration increases from the MDL, hence the MDL determined under these circumstances may not truly reflect method variance at lower analyte concentrations.

4. (a) Take a minimum of seven aliquots of the sample to be used to calculate the method detection limit and process each through the entire analytical method. Make all computations according to the defined method with final results in the method reporting units. If a blank measurement is required to calculate the measured level of analyte, obtain a separate blank measurement for each sample aliquot analyzed. The average blank measurement is subtracted from the respective sample measurements.

(b) It may be economically and technically desirable to evaluate the estimated method detection limit before proceeding with 4a. This will: (1) Prevent repeating this entire procedure when the costs of analyses are high and (2) insure that the procedure is being conducted at the correct concentration. It is quite possible that an inflated MDL will be calculated from data obtained at many times the real MDL even though the level of analyte is less than five times the calculated method detection limit. To insure that the estimate of the method detection limit is a good estimate, it is necessary to determine that a lower concentration of analyte will not result in a significantly lower method detection limit. Take two aliquots of the sample to be used to calculate the method detection limit and process each through the entire method, including blank measurements as described above in 4a. Evaluate these data:

(1) If these measurements indicate the sample is in desirable range for determination of the MDL, take five additional aliquots and proceed. Use all seven measurements for calculation of the MDL.

(2) If these measurements indicate the sample is not in correct range, reestimate the MDL, obtain new sample as in 3 and repeat either 4a or 4b.

5. Calculate the variance ($S^2$) and standard deviation ($S$) of the replicate measurements, as follows:

$$
S^2 = \frac{1}{n-1} \left[ \sum_{i=1}^{n} x_i^2 - \left( \sum_{i=1}^{n} x_i \right)^2 \right]
$$

where:

$$
S = (S^2)^{1/2}
$$

$X_i$: i=1 to n, are the analytical results in the final method reporting units obtained from
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the n sample aliquots and Σ refers to the sum of the X values from i=1 to n.
6. (a) Compute the MDL as follows:

\[ \text{MDL} = T_{(n-1,1-\alpha=0.99)}(S) \]

where:
\[ \text{MDL} \] = the method detection limit
\[ T_{(n-1,1-\alpha=0.99)} \] = the student's t value appropriate for a 99% confidence level and a standard deviation estimate with n-1 degrees of freedom. See Table.
\[ S \] = standard deviation of the replicate analyses.

(b) The 95% confidence interval estimates for the MDL derived in 6a are computed according to the following equations derived from percentiles of the chi square over degrees of freedom distribution (\( \chi^2/df \)).

\[ \text{LCL} = 0.64 \times \text{MDL} \]
\[ \text{UCL} = 2.20 \times \text{MDL} \] where: LCL and UCL are the lower and upper 95% confidence limits respectively based on seven aliquots.

7. Optional iterative procedure to verify the reasonableness of the estimate of the MDL and subsequent MDL determinations.

(a) If this is the initial attempt to compute MDL based on the estimate of MDL formulated in Step 1, take the MDL as calculated in Step 6, spike the matrix at this calculated MDL and proceed through the procedure starting with Step 4.

(b) If this is the second or later iteration of the MDL calculation, use \( S^2 \) from the current MDL calculation and \( S^2 \) from the previous MDL calculation to compute the F-ratio. The F-ratio is calculated by substituting the larger \( S^2 \) into the numerator \( S^2_a \) and the other into the denominator \( S^2_p \). The computed F-ratio is then compared with the F-ratio found in the table which is 3.05 as follows: if \( S^2_a/S^2_p < 3.05 \), then compute the pooled standard deviation by the following equation:

\[ S_{\text{pooled}} = \left( \frac{6S^2_a + 6S^2_p}{12} \right)^{\frac{1}{2}} \]

if \( S^2_a/S^2_p > 3.05 \), respike at the most recent calculated MDL and process the samples through the procedure starting with Step 4. If the most recent calculated MDL does not permit qualitative identification when samples are spiked at that level, report the MDL as a concentration over degrees of freedom distribution.

\[ \text{LCL} = 0.72 \times \text{MDL} \]
\[ \text{UCL} = 1.65 \times \text{MDL} \] where LCL and UCL are the lower and upper 95% confidence limits respectively based on 14 aliquots.

### TABLES OF STUDENTS’ T VALUES AT THE 99 PERCENT CONFIDENCE LEVEL—Continued

<table>
<thead>
<tr>
<th>Number of replicates</th>
<th>Degrees of freedom (n-1)</th>
<th>( t_{(n-1,\alpha)} )</th>
</tr>
</thead>
<tbody>
<tr>
<td>7</td>
<td>6</td>
<td>3.143</td>
</tr>
</tbody>
</table>

### Reporting

The analytical method used must be specifically identified by number or title ald the MDL for each analyte expressed in the appropriate method reporting units. If the analytical method permits options which affect the method detection limit, these conditions must be specified with the MDL value. The sample matrix used to determine the MDL must also be identified with MDL value. Report the mean analyte level with the MDL and indicate if the MDL procedure was iterated. If a laboratory standard or a sample that contained a known amount analyte was used for this determination, also report the mean recovery.
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If the level of analyte in the sample was below the determined MDL or exceeds 10 times the MDL of the analyte in reagent water, do not report a value for the MDL.


APPENDIX C TO PART 136—INDUCTIVELY COUPLED PLASMA—ATOMIC EMISSION SPECTROMETRIC METHOD FOR TRACE ELEMENT ANALYSIS OF WATER AND WASTES METHOD 200.7

1. Scope and Application

1.1 This method may be used for the determination of dissolved, suspended, or total elements in drinking water, surface water, and domestic and industrial wastewaters.

1.2 Dissolved elements are determined in filtered and acidified samples. Appropriate steps must be taken in all analyses to ensure that potential interferences are taken into account. This is especially true when dissolved solids exceed 1500 mg/L. (See Section 5.)

1.3 Total elements are determined after appropriate digestion procedures are performed. Since digestion techniques increase the dissolved solids content of the samples, appropriate steps must be taken to correct for potential interference effects. (See Section 5.)

1.4 Table 1 lists elements for which this method applies along with recommended wavelengths and typical estimated instrumental detection limits using conventional pneumatic nebulization. Actual working detection limits are sample dependent and as the sample matrix varies, these concentrations may also vary. In time, other elements may be added as more information becomes available and as required.

1.5 Because of the differences between various makes and models of satisfactory instruments, no detailed instrumental operating instructions can be provided. Instead, the analyst is referred to the instruction provided by the manufacturer of the particular instrument.

2. Summary of Method

2.1 The method describes a technique for the simultaneous or sequential multielement determination of trace elements in solution. The basis of the method is the measurement of atomic emission by an optical spectroscopic technique. Samples are nebulized and the aerosol that is produced is transported to the plasma torch where excitation occurs. Characteristic atomic-line emission spectra are produced by a radio-frequency inductively coupled plasma (ICP). The spectra are dispersed by a grating spectrometer and the intensities of the lines are monitored by photomultiplier tubes. The photocurrents from the photomultiplier tubes are processed and controlled by a computer system. A background correction technique is required to compensate for variable background contribution to the determination of trace elements. Background must be measured adjacent to analyte lines on samples during analysis. The position selected for the background intensity measurement, on either or both sides of the analytical line, will be determined by the complexity of the spectrum adjacent to the analyte line. The position used must be free of spectral interference and reflect the same change in background intensity as occurs at the analyte wavelength measured. Background correction is not required in cases of line broadening where a background correction measurement would actually degrade the analytical result. The possibility of additional interferences named in 5.1 (and tests for their presence as described in 5.2) should also be recognized and appropriate corrections made.

3. Definitions

3.1 Dissolved—Those elements which will pass through a 0.45 μm membrane filter.

3.2 Suspended—Those elements which are retained by a 0.45 μm membrane filter.

3.3 Total—The concentration determined on an unfiltered sample following vigorous digestion (Section 9.3), or the sum of the dissolved plus suspended concentrations. (Section 9.1 plus 9.2.)

3.4 Total recoverable—The concentration determined on an unfiltered sample following treatment with hot, dilute mineral acid (Section 9.4).

3.5 Instrumental detection limit—The concentration equivalent to a signal, due to the analyte, which is equal to three times the standard deviation of a series of ten replicate measurements of a reagent blank signal at the same wavelength.

3.6 Sensitivity—The slope of the analytical curve, i.e., functional relationship between emission intensity and concentration.

3.7 Instrument check standard—A multielement standard of known concentrations prepared by the analyst to monitor and verify instrument performance on a daily basis. (See 7.6.1)

3.8 Interference check sample—A solution containing both interfering and analyte elements of known concentration that can be used to verify background and interelement correction factors. (See 7.6.2.)

3.9 Quality control sample—A solution obtained from an outside source having known, concentration values to be used to verify the instrumental detection limit (Section 9.1).

3.10 Calibration standards—A series of known standard solutions used by the analyst for calibration of the instrument (i.e., preparation of the analytical curve). (See 7.4)