§ 493.1261 Standard: Bacteriology.
(a) The laboratory must check the following for positive and negative reactivity using control organisms:
(1) Each day of use for beta-lactamase methods other than CefinaseTM.
(2) Each week of use for Gram stains.
(3) When each batch (prepared in-house), lot number (commercially prepared), and shipment of antisera is prepared or opened, and once every 6 months thereafter.
(b) For antimicrobial susceptibility tests, the laboratory must check each batch of media and each lot number and shipment of antimicrobial agent(s) before, or concurrent with, initial use, using approved control organisms.
(1) Each day tests are performed, the laboratory must use the appropriate control organism(s) to check the procedure.
(2) The laboratory’s zone sizes or minimum inhibitory concentration for control organisms must be within established limits before reporting patient results.
(c) The laboratory must document all control procedures performed, as specified in this section.

§ 493.1262 Standard: Mycobacteriology.
(a) Each day of use, the laboratory must check all reagents or test procedures used for mycobacteria identification with at least one acid-fast organism that produces a positive reaction and an acid-fast organism that produces a negative reaction.
(b) For antimycobacterial susceptibility tests, the laboratory must check each batch of media and each lot number and shipment of antituberculosis agent(s) before, or concurrent with, initial use, using an appropriate control organism(s).
(1) The laboratory must establish limits for acceptable control results.
(2) Each week tests are performed, the laboratory must use the appropriate control organism(s) to check the procedure.
(3) The results for the control organism(s) must be within established limits before reporting patient results.
(c) The laboratory must document all control procedures performed, as specified in this section.

§ 493.1263 Standard: Parasitology.
(a) The laboratory must have available a reference collection of slides or photographs and, if available, gross specimens for identification of parasites and use these references in the laboratory for appropriate comparison with diagnostic specimens.
(b) The laboratory must calibrate and use the calibrated ocular micrometer for determining the size of ova and parasites, if size is a critical parameter.
(c) Each month of use, the laboratory must check permanent stains using a fecal sample control material that will demonstrate staining characteristics.